

Selective ATP-Competitive Inhibitors of TOR Suppress Rapamycin-Insensitive Function of TORC2 in *Saccharomyces cerevisiae*

Qingsong Liu,^{†,‡,▽} Tao Ren,^{§,▽} Tara Fresques,^{||,▽} Wolfgang Oppliger,[⊥] Brad J. Niles,^{||} Wooyoung Hur,^{†,‡} David M. Sabatini,^{#,||,○} Michael N. Hall,[⊥] Ted Powers,^{||} and Nathanael S. Gray^{*,†,‡}

[†]Department of Cancer Biology, Dana-Farber Cancer Institute, 44 Binney Street, Boston, Massachusetts 02115, United States

[‡]Department of Biology Chemistry and Molecular Pharmacology, Harvard Medical School, 250 Longwood Avenue, Boston, Massachusetts 02115, United States

[§]Harvard Medical School, Boston, Massachusetts 02115, United States

^{||}Department of Molecular and Cellular Biology, College of Biological Sciences, University of California, Davis, California 95616, United States

[⊥]Biozentrum, University of Basel, CH-4056 Basel, Switzerland

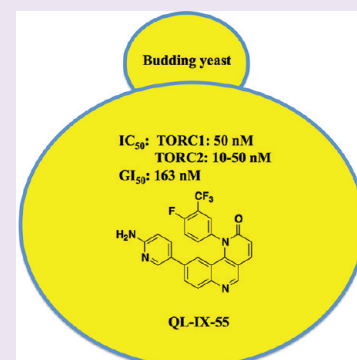
[#]Whitehead Institute for Biomedical Research, 9 Cambridge Center, Cambridge, Massachusetts 02142, United States

^{||}Howard Hughes Medical Institute, Department of Biology, Massachusetts Institute of Technology, Cambridge, Massachusetts 02139, United States

[○]Koch Center for Integrative Cancer Research at MIT, 77 Massachusetts Avenue, Cambridge, Massachusetts 02139, United States

Supporting Information

ABSTRACT: The target of rapamycin (TOR) is a critical regulator of growth, survival, and energy metabolism. The allosteric TORC1 inhibitor rapamycin has been used extensively to elucidate the TOR related signal pathway but is limited by its inability to inhibit TORC2. We used an unbiased cell proliferation assay of a kinase inhibitor library to discover QL-IX-55 as a potent inhibitor of *S. cerevisiae* growth. The functional target of QL-IX-55 is the ATP-binding site of TOR2 as evidenced by the discovery of resistant alleles of TOR2 through rational design and unbiased selection strategies. QL-IX-55 is capable of potently inhibiting both TOR complex 1 and 2 (TORC1 and TORC2) as demonstrated by biochemical IP kinase assays ($IC_{50} < 50$ nM) and cellular assays for inhibition of substrate YPK1 phosphorylation. In contrast to rapamycin, QL-IX-55 is capable of inhibiting TORC2-dependent transcription, which suggests that this compound will be a powerful probe to dissect the Tor2/TORC2-related signaling pathway in yeast.



The resurgence of pathogenic yeast infections has stimulated interest in developing new classes of antifungal agents.¹ Currently approved antifungals target a limited number of biological pathways that are unique to fungi in order to obtain agents that are well-tolerated in humans. Protein kinases have become an intensely pursued enzyme class particularly for the treatment of cancer where 12 compounds have received approval and approximately 120 compounds are in different stages of clinical testing.² The kinome of *Saccharomyces cerevisiae* encodes approximately 130 protein kinases of which at least 15 have been shown to be essential based upon genetic deletion.³ The analogue-sensitive (AS) approach to generating mutant kinases that can be uniquely and potently inhibited by small molecules such as 1-naphthylmethyl-PP1 (1-NM-PP1) has provided proof that pharmacological inhibition of kinases such as Cdc28, Pho85, Hog1, and others can lead to cytostatic or cytotoxic effects in *S. cerevisiae*.⁴⁻⁶ Recently a small molecule TORC1 inhibitor, CID3528206, which is believed to act by a mechanism similar to that of rapamycin, was discovered using a multiplex flow cytometry assay.⁷ We sought to address whether

the large pharmacopeia of small molecule ATP-competitive inhibitors that have been developed to target human kinases could be leveraged to discover inhibitors of homologous or non-homologous yeast kinases that are efficacious in cellular assays. Should potent inhibitors of *S. cerevisiae* kinases be discovered, they will likely need to be optimized to evade the efficient efflux mechanisms of yeast, to possess selectivity relative to human kinases, and to have the ability to overcome resistance that is likely to result from emergence of point mutations in target kinases

To discover efficacious *in vivo* inhibitors of yeast kinases, we performed a medium-throughput growth assay of a 1000-compound kinase library containing most of the known ATP-competitive pharmacophores and a large number of clinical stage kinase inhibitors. As our first question was how many of these compounds would recognize yeast kinases and lead to

Received: February 10, 2012

Accepted: April 11, 2012

Published: April 11, 2012

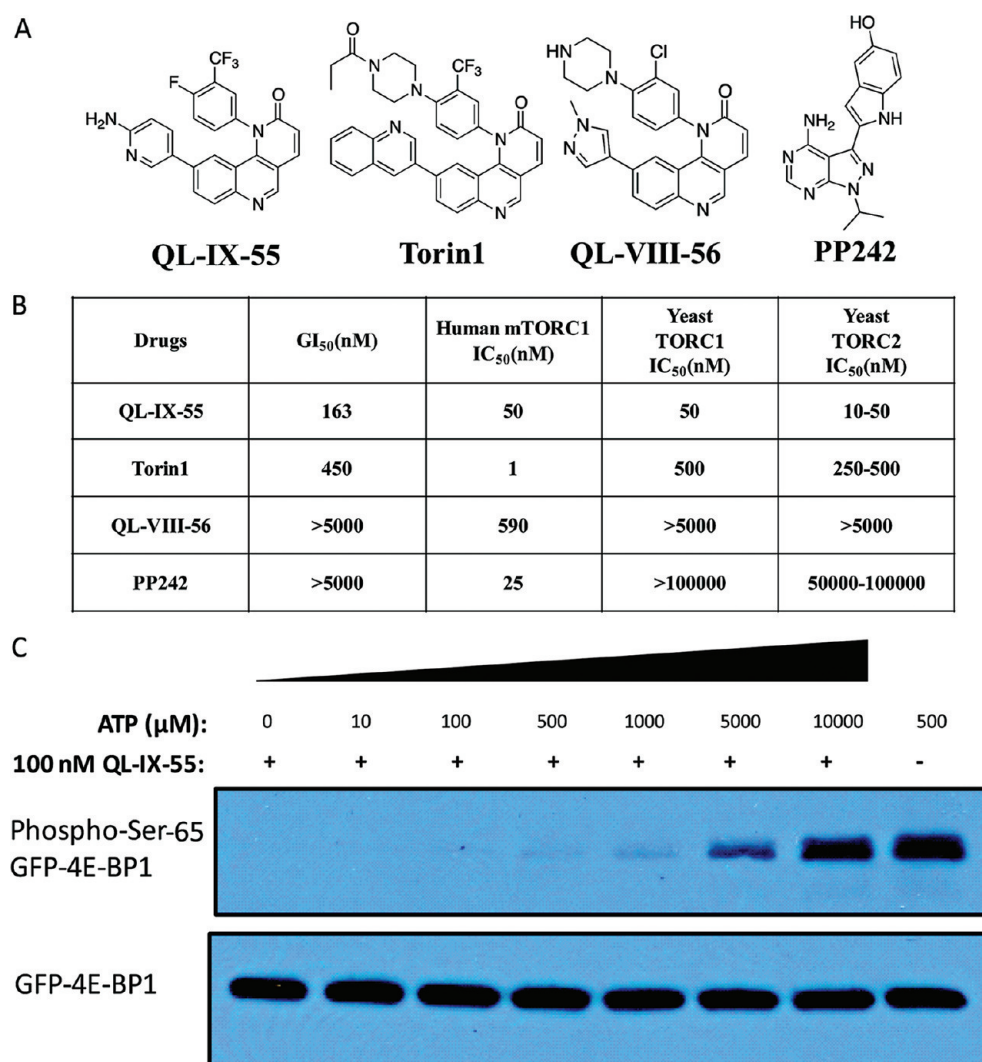


Figure 1. Discovery and characterization of QL-IX-55 as a TORC1/2 inhibitor. (A) Chemical structures of QL-IX-55, Torin 1, QL-VIII-56, and PP242. (B) QL-IX-55 inhibits TORC1 and TORC2. (C) QL-IX-55 is an ATP competitive inhibitor.

growth inhibition, we decided to circumvent the drug penetration and efflux problem by using a “drug-sensitized” yeast strain. We used the RDY98 yeast strain, which contains deletion ERG6, an essential enzyme in ergosterol biosynthesis and in PDR1 and PDR3, two ABC-multidrug transporters^{8,9} (Supplementary Figure S1). This strain has previously been shown to greatly facilitate uptake of a variety of different drugs in yeast. Screening at a concentration of 10 μM in liquid phase cultures, we discovered that a number of analogues of Torin1, an inhibitor that we had previously developed as the first ATP-competitive inhibitor of mTOR, were strongly growth-inhibitory.¹⁰ Follow-up titrations of the most active compound resulted in the identification of QL-IX-55 as the most potent Torin1 analogue, which inhibited growth to 50% of the DMSO control at a concentration of 163 nM (Figure 1A and Supplementary Figure S2). QL-IX-55 is not capable of inhibiting growth of wild-type yeast at concentrations below 10 μM, demonstrating that the drug-sensitizing mutations are required for its cellular efficacy (data not shown). Interestingly, other reported potent inhibitors of human mTOR that were present in our library, such as AZD8055, KU63794, WYE354, and PP242, were not growth-inhibitory up to a concentration of 1 μM (Supplementary Table S1).^{11–14}

We next sought to identify what molecular target(s) might be responsible for the growth-inhibitory effects of QL-IX-55. Because QL-IX-55 is a structural analogue of Torin1, we naturally speculated that QL-IX-55 might also target TOR in yeast. Yeast TOR was originally identified as the pharmacological target of the natural product rapamycin through genetic resistance screens and affinity chromatography approaches.¹⁵ Yeast TOR exhibits a high degree of sequence and domain conservation relative to human TOR and rapamycin, which binds to the FRB domain, and is known to be a potent allosteric inhibitor of the kinase activity of both human and yeast enzymes. The kinase domains of *S. cerevisiae* TOR and human mTOR bear over 60% sequence identity in the catalytic domain, and there are substitutions of approximately 10 amino acids that are predicted to form the ATP-binding pocket (Supplementary Figure S3). However, unlike in mammalian cells, which encode only a single TOR gene, in yeast there are two TOR genes, TOR1 and TOR2.^{16,17} The TOR1 and TOR2 proteins form two functionally and structurally distinct complexes termed TORC1 and TORC2. Yeast TORC1 consists of Kog1, Tco89, Lst8, and either TOR1 or TOR2. In contrast, TORC2 contains only TOR2 in complex with Avo1, Avo2, Avo3, Bit61, and Lst8.¹⁸ To determine whether

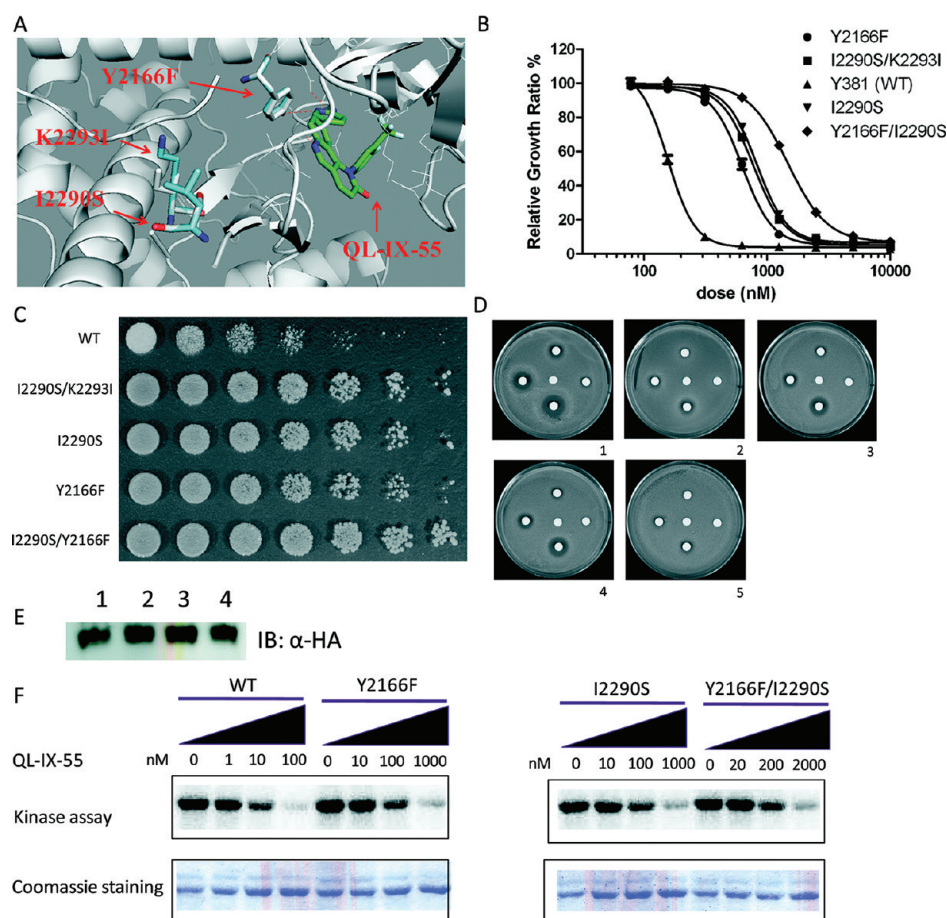


Figure 2. QL-IX-55 inhibition of TOR2. (A) Docking QL-IX-55 to a mTOR homology model. (B–D) Growth inhibition assay, Clonogenic assay, and Halo assay of QL-IX-55 on TOR2 wide/mutant strains. (E) Detection of HA-tagged Tor2 (HA-TOR2) by immunoblotting with Anti-HA antibody. (F) *In vitro* TOR2 kinase assay.

QL-IX-55 inhibits TORC1 and/or TORC2, we performed kinase assays with both complexes individually immunoprecipitated from yeast cells expressing tagged Kog1 (TORC1) or Avo3 (TORC2).¹⁸ QL-IX-55 potently inhibited both complexes with an IC_{50} of approximately 50 nM (Figure 1B). Torin1, which exhibits a less potent GI_{50} of 450 nM, is also a less potent inhibitor of TORC1 (IC_{50} = 500 nM) and of TORC2 (IC_{50} = 250–500 nM). A close structural analogue, QL-VIII-56, which was inactive as a growth inhibitor below concentrations of 10 μ M, also lost its ability to inhibit TORC1 and TORC2. To elucidate the mechanism of the inhibition, we performed an ATP competition assay, which demonstrated that QL-IX-55 is an ATP-competitive inhibitor of TOR2/TORC2. This mode of inhibition is fundamentally distinct from that of rapamycin, which is non-ATP-competitive and binds to the FRB domain located N-terminal to the kinase domain. (Figure 1C). Since there are no readily available methods to screen for selectivity comprehensively against all yeast kinases, we instead profiled QL-IX-55 (1 μ M) against a panel of 440 human kinases using the DiscoverX KinomeScan technology (Supplementary Table S2). This analysis demonstrated that QL-IX-55 maintained the ability to bind to human mTOR and structurally related PI3K but is otherwise a remarkably selective kinase inhibitor, comparable to some of the most highly selective ATP-competitive inhibitors currently known.

To determine whether inhibition of TOR1 and/or TOR2 kinase activity was functionally related to the potent growth-

inhibitory activity of QL-IX-55, we performed both molecular model-guided mutagenesis and an unbiased drug selection screen. A molecular model for QL-IX-55 in complex with yeast TOR2 was constructed based upon the most closely related crystal structure (PI3K γ , PDB id: 3DBS). This model predicted the existence of a hydrogen bond between the phenol of Y2166 located deep in the ATP-binding site with the aminopyrimidine motif of the inhibitor (Figure 2A). To evaluate this hypothetical interaction, we created a Y2166F mutation in TOR2 and constructed a yeast strain bearing this mutation integrated into the native locus using homologous recombination. We also performed an unbiased selection experiment where yeast cells were grown for 3 days in rich medium containing 300 nM QL-IX-55. Single clones were isolated by streaking the selected cultures onto YPD agar plates also containing 300 nM QL-IX-55. Targeted sequencing of TOR1 and TOR2 from genomic DNA isolated from resistant colonies revealed no mutations in TOR1 and a recurrent I2290S/K2293I mutation in TOR2. Interestingly, these mutations map closely to the ATP-binding site, and I2290S is located four amino acids N-terminal to the highly conserved DFG motif, which marks the start of the activation loop and often undergoes dramatic rearrangements upon inhibitor binding. As other undetected mutations may also be present in these resistant colonies, we re-engineered strains with I2290S, K2293I, I2290S/K2293I, and I2290S/Y2166F TOR2 alleles and examined their ability to grow in the presence of QL-IX-55. In liquid-phase cultures, the I2290S

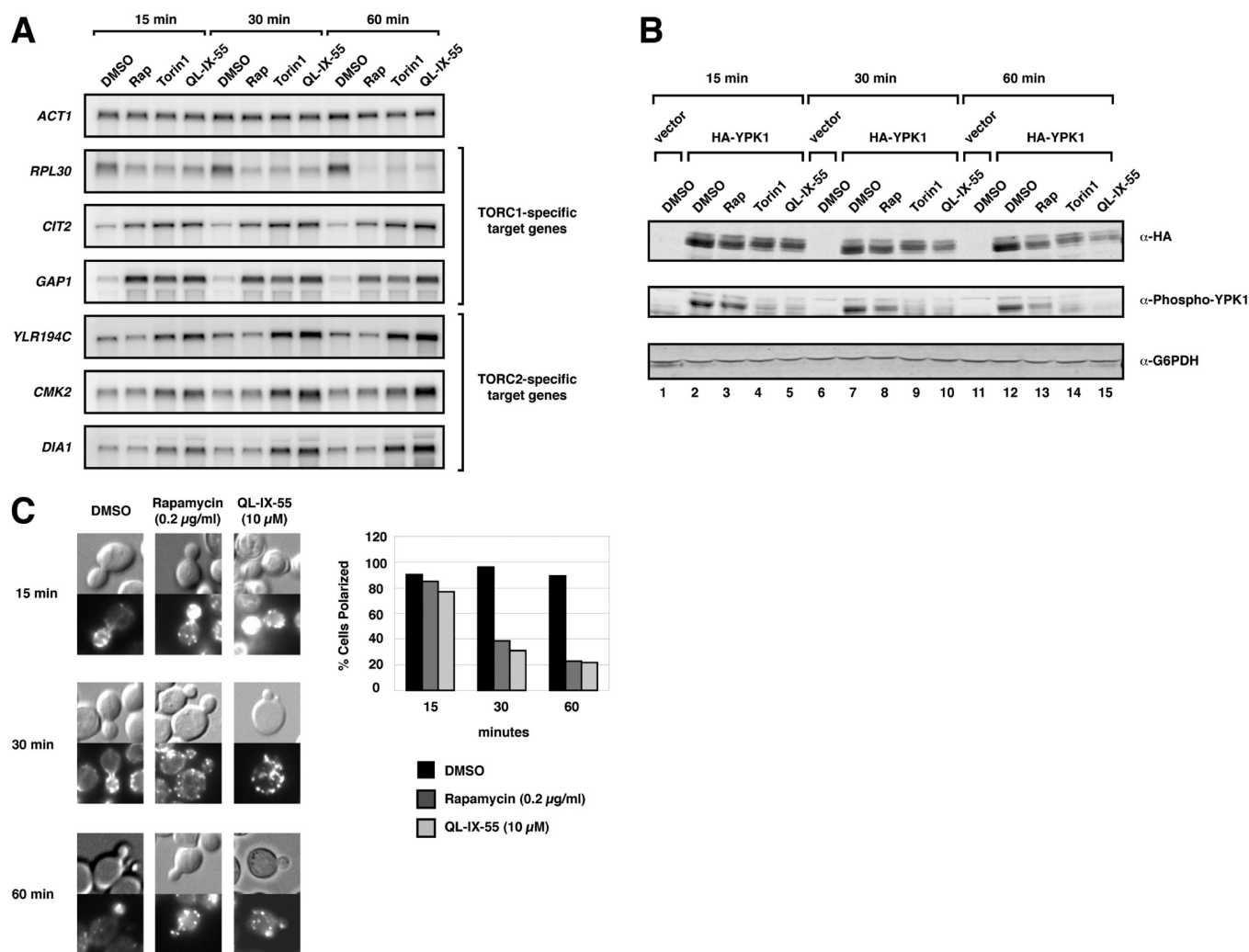


Figure 3. QL-IX-55 inhibition of both TORC1 and TORC2 signaling. (A) Northern blot analysis of total mRNA level. (B) TORC2-dependent phosphorylation of YPK1 inhibition. (C) Actin cytoskeletal organization perturbation.

mutation resulted in a 5-fold increase in GI_{50} , the Y2166F mutation induced a 4-fold increase in GI_{50} , the K2293I mutation had no effect, and the double I2290S/Y2166F double mutation had an additive effect that resulted in a 9-fold elevation in GI_{50} relative to wild-type (Figure 2B and Supplementary Table S3). A similar resistance profile was observed when the assays were repeated using colony dilution and halo assays (Figure 2C,D). To determine whether growth resistance correlated with a loss in ability of QL-IX-55 to inhibit the kinase activity of mutant TOR2, we performed immunoprecipitation assays from I2290S/Y2166F TOR2 yeast. The IC_{50} for inhibition of I2290S/Y2166F TOR2 was elevated approximately 20-fold relative to that of wild-type TOR2, which is consistent with TOR being a functionally relevant target for the growth-inhibitory phenotype (Figure 2E,F).

We next sought to use our novel inhibitor to investigate the physiological consequences of ATP-competitive inhibition of TOR kinase activity, in comparison to allosteric inhibition using rapamycin. In yeast, TORC1 but not TORC2 is sensitive to rapamycin.¹⁷ By looking at gene expression, we discovered that QL-IX-55 was capable of modulating transcriptional outputs of both TORC1 and TORC2. For example, similar to rapamycin, QL-IX-55 decreased expression of RPL30 and increased

expression of CIT2 and GAP1, as demonstrated by Northern blot analysis (Figure 3A).¹⁹ However, unlike rapamycin, QL-IX-55 also modulated transcriptional outputs specific for TORC2, including increased expression of YLR194C, CMK2, and DIA1 (Figure 3A).²⁰ We also observed that QL-IX-55, but not rapamycin, rapidly inhibited TORC2-dependent phosphorylation of Ypk1, a recently identified substrate of TORC2 (Figure 3B, middle panel, compare lanes 3 and 5). We next investigated the effects of QL-IX-55 on actin morphology, where inhibition of either TORC1 or TORC2 is known to result in depolarization of actin patch structures.²¹ Here we observed that both rapamycin and QL-IX-55 perturbed actin morphology with similar kinetics, where fewer than 20% of cells displayed normal polarization following 60 min of drug treatment (Figure 3C).

The TOR (Target of Rapamycin) signaling network couples cell growth to intracellular and environmental cues in all eukaryotic organisms examined to date.²² This network was discovered through the action of rapamycin, an immunosuppressant and antifungal compound that acts as a specific allosteric inhibitor of the TOR kinase, a member of the PI3-like kinase family of serine/threonine protein kinases.¹⁵ TOR associates with a number of other proteins to form two distinct protein complexes, termed TORC1 and TORC2 (or mTORC1

and mTORC2 in mammalian cells), where TORC1 is uniquely inhibited by rapamycin. These complexes control diverse downstream processes, including protein synthesis and actin cytoskeletal organization, and thus collaboratively regulate both temporal and spatial aspects of cell growth. Much of what has been learned about TOR signaling has come from studies using budding yeast, *S. cerevisiae*, and this remains an important model system for exploring the architecture and function of TOR. In particular, rapamycin has been invaluable as a specific small molecule inhibitor of TOR activity and has revealed a role for TOR signaling in a wide array of activities, including protein synthesis, ribosome biogenesis, autophagy, nutrient-regulated gene expression, and protein trafficking.²² Because rapamycin targets only TORC1, however, our understanding of the scope of processes controlled by TORC2 has been limited by comparison. To a large extent this problem has now been circumvented in mammalian cells by the development of a cadre of ATP-competitive inhibitors specific for mTOR, which have been shown to inhibit the activity of both mTORC1 and mTORC2.²³ Unfortunately these compounds have proven to be mostly ineffective against yeast, either because of problems with drug uptake and/or retention, subtle differences in the structure of the ATP binding pocket between yeast TOR and mTOR, or both.

By screening a diverse library of 1000 different kinase inhibitors for inhibitors of yeast growth, we have discovered the compound QL-IX-55, a structural analogue of Torin1, which functions as an ATP-competitive inhibitor of yeast TOR and is capable of potentially inhibiting the kinase activity and modulating the outputs of both TORC1 and TORC2. As QL-IX-55 inhibits both TORC1 and TORC2, it most likely targets both TOR1 and TOR2. QL-IX-55 resistance mutations were obtained only in *TOR2* because only TOR2 is in both TORC1 and TORC2 and mutations in *TOR1* would not affect the sensitivity of TORC2. The ability of QL-IX-55 to potentially inhibit yeast growth depends on its ability to inhibit TOR *in vivo*. However, our currently best *TOR2* resistance allele I2290S/Y2166F confers approximately 20-fold resistance using a *TOR2 in vitro* kinase assay but only 9-fold resistance to growth inhibition. This apparent discrepancy is likely due to QL-IX-55 also targeting TOR1 and possibly other yeast enzymes.

Just as rapamycin has proven to be invaluable for mapping TORC1-dependent outputs, QL-IX-55 will serve as a useful means to map both TORC1- and TORC2-dependent outputs in yeast. Our study also demonstrates that ATP-competitive inhibition of TOR is a pharmacologically valid means of obtaining antifungal activity. QL-IX-55 possesses two major deficits that must be overcome to improve its use as a pharmacological tool and potential as a therapeutic lead compound. First, the ability of QL-IX-55 to accumulate in yeast needs to be improved to obtain activity against wild-type and pathogenic yeast. Second, although QL-IX-55 is approximately a 50-fold weaker inhibitor of human TORC1 and a 10-fold more potent inhibitor of yeast TORC1 relative to Torin1, further optimization of its yeast versus human selectivity will be needed. Our screen also demonstrated that highly potent ATP-competitive human mTOR inhibitors such as AZD8055, KU63794, WYE354, and PP242 are not growth-inhibitory up to a concentration of 1 μ M, which again demonstrates that achieving selectivity between yeast and human mTOR can be accomplished. Finally, our screen demonstrated that surprisingly few of the currently known human kinase inhibitors

including highly promiscuous compounds such as dasatinib, sunitinib, and sorafenib possess any significant antifungal activity even at a high concentration of 10 μ M. This gives credence to the notion that achieving selectivity for fungal *versus* human kinases is an achievable goal but will require dedicated medicinal chemistry efforts potentially with new chemical templates.

METHODS

Drugs, yeast strains, and antibodies are described in detail in the Supporting Information.

Drug-Induced Mutagenesis and Mutant Regeneration.

RDY98 yeast cells were grown in YPD liquid medium to mid-log phase and then diluted to $A_{600} = 0.05$ using fresh medium containing 200 nM QL-IX-55 for 2 days incubation. A couple of cell divisions later, 200 μ L samples of cultures were spread on YPD agar plates containing 300 nM QL-IX-55 for 3 days until colonies appear. Seventy-two colonies were picked and streaked on YPD agar plates containing 300 nM QL-IX-55. TOR1 and TOR2 genes were amplified and sequenced. Homologous recombination was used for generation of yeast mutants.¹ The LiAc method was applied for yeast transformation.

In Vitro TORC1/2 Biochemical Assay. TORC1 and TORC2 were purified by Protein A Sepharose cross-linked with mouse monoclonal HA antibody from exponentially growing *KOG1-HA* and *AVO3-HA* cells, respectively. Inhibitors were tested using 8-fold serial dilutions from 100 μ M to 10 nM. Kinase reactions were performed in kinase buffer in a final volume of 30 μ L containing 20 μ g of PHAS-I (4EBP1). Kinase reactions were started with the addition of 80 μ M ATP and 5 μ Ci [γ -³²P]ATP. Twenty-three microliters of the supernatants were transferred to Filter P18 (Whatman). The incorporation of ³²P was measured by liquid-scintillation analyzer. IC₅₀ values were determined by fitting the data to a sigmoid dose-response curve using the BioDataFit 1.02 software package.

Other methods are described in detail in the Supplemental Experimental Procedures.

ASSOCIATED CONTENT

Supporting Information

Supplemental experimental procedures, Tables S1–S3, Figures S1–S3, and supplemental references. This material is available free of charge *via* the Internet at <http://pubs.acs.org>.

AUTHOR INFORMATION

Corresponding Author

*E-mail: Nathanael_Gray@dfci.harvard.edu.

Author Contributions

These authors contributed equally to this work.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We thank K. Struhl (Harvard Medical School) for providing yeast plasmids and medium and H. H. Wong (Harvard Medical School) for productive discussion. We thank T. Kapoor for suggesting we perform an unbiased kinase library screen for antifungals. We are also acknowledge a NERCE Grant (U54 A1057159) for partial financial support to R.T., a NIH LINC Grant (HG006097) for the support of Q.L., and a NIH Grant (R01GM86387) to T.P.

REFERENCES

(1) Arendrup, M. C., Fisher, B. T., and Zaoutis, T. E. (2009) Invasive fungal infections in the paediatric and neonatal population: diagnostics and management issues. *Clin. Microbiol. Infect.* 15, 613–624.

- (2) Maggon, K. (2011) *Tyrosine Kinase Inhibitors R&D Review*, 49.
- (3) Ptacek, J., Devgan, G., Michaud, G., Zhu, H., Zhu, X., Fasolo, J., Guo, H., Jona, G., Breitskreutz, A., Sopko, R., McCartney, R. R., Schmidt, M. C., Rachidi, N., Lee, S. J., Mah, A. S., Meng, L., Stark, M. J., Stern, D. F., DeVirgilio, C., Tyers, M., Andrews, B., Gerstein, M., Schweitzer, B., Predki, P. F., and Snyder, M. (2005) Global analysis of protein phosphorylation in yeast. *Nature* 438, 679–684.
- (4) Ira, G., Pelliccioli, A., Balijja, A., Wang, X., Fiorani, S., Carotenuto, W., Liberi, G., Bressan, D., Wan, L., Hollingsworth, N. M., Haber, J. E., and Foiani, M. (2004) DNA end resection, homologous recombination and DNA damage checkpoint activation require CDK1. *Nature* 431, 1011–1017.
- (5) Dinér, P., Veide, Vilg. J., Kjellén, J., Migdal, I., Andersson, T., Gebbia, M., Giaever, G., Nislow, C., Hohmann, S., Wysocki, R., Tamás, M. J., and Grøtli, M. (2011) Design, synthesis, and characterization of a highly effective Hog1 inhibitor: a powerful tool for analyzing MAP kinase signaling in yeast. *PLoS One* 6, e20012.
- (6) Sussman, A., Huss, K., Chio, L. C., Heidler, S., Shaw, M., Ma, D., Zhu, G., Campbell, R. M., Park, T. S., Kulanthaivel, P., Scott, J. E., Carpenter, J. W., Strega, M. A., Belvo, M. D., Swartling, J. R., Fischl, A., Yeh, W. K., Shih, C., and Ye, X. S. (2004) Discovery of cercosporamide, a known antifungal natural product, as a selective Pkc1 kinase inhibitor through high-throughput screening. *Eukaryot. Cell* 3, 932–943.
- (7) Chen, J., Young, S. M., Allen, C., Seeber, A., Péli-Gulli, M. P., Panchaud, N., Waller, A., Ursu, O., Yao, T., Golden, J. E., Strouse, J. J., Carter, M. B., Kang, H., Bologa, C. G., Foutz, T. D., Edwards, B. S., Peterson, B. R., Aubé, J., Werner-Washburne, M., Loewith, R. J., DeVirgilio, C., and Sklar, L. A. (2012) Identification of a small molecule yeast TORC1 inhibitor with a multiplex screen based on flow cytometry. *ACS Chem Biol.* DOI: 10.1021/cb200452r.
- (8) Gaber, R. F., Copple, D. M., Kennedy, B. K., Vidal, M., and Bard, M. (1989) The yeast gene ERG6 is required for normal membrane function but is not essential for biosynthesis of the cell-cycle-sparking sterol. *Mol. Cell. Biol.* 9, 3447–3456.
- (9) Nawrocki, A., Fey, S. J., Goffeau, A., Roepstorff, P., and Larsen, P. M. (2001) The effects of transcription regulating genes PDR1, pdr1–3 and PDR3 in pleiotropic drug resistance. *Proteomics* 1, 1022–1032.
- (10) Liu, Q., Chang, J. W., Wang, J., Kang, S. A., Thoreen, C. C., Markhard, A., Hur, W., Zhang, J., Sim, T., Sabatini, D. M., and Gray, N. S. (2010) Discovery of 1-(4-(4-propionylpiperazin-1-yl)-3-(trifluoromethyl)phenyl)-9-(quinolin-3-yl)benzo[h][1,6]naphthyridin-2(1H)-one as a highly potent, selective mammalian target of rapamycin (mTOR) inhibitor for the treatment of cancer. *J. Med. Chem.* 53, 7146–7155.
- (11) Chresta, C. M., Davies, B. R., Hickson, I., Harding, T., Cosulich, S., Critchlow, S. E., Vincent, J. P., Ellston, R., Jones, D., Sini, P., James, D., Howard, Z., Dudley, P., Hughes, G., Smith, L., Maguire, S., Hummersone, M., Malagu, K., Menear, K., Jenkins, R., Jacobsen, M., Smith, G. C., Guichard, S., and Pass, M. (2010) AZD8055 is a potent, selective, and orally bioavailable ATP-competitive mammalian target of rapamycin kinase inhibitor with in vitro and in vivo antitumor activity. *Cancer Res.* 70, 288–298.
- (12) García-Martínez, J. M., Moran, J., Clarke, R. G., Gray, A., Cosulich, S. C., Chresta, C. M., and Alessi, D. R. (2009) Ku-0063794 is a specific inhibitor of the mammalian target of rapamycin (mTOR). *Biochem. J.* 421, 29–42.
- (13) Yu, K., Toral-Barza, L., Shi, C., Zhang, W. G., Lucas, J., Shor, B., Kim, J., Verheijen, J., Curran, K., Malwitz, D. J., Cole, D. C., Ellingboe, J., Ayrál-Kaloustian, S., Mansour, T. S., Gibbons, J. J., Abraham, R. T., Nowak, P., and Zask, A. (2009) Biochemical, cellular, and in vivo activity of novel ATP-competitive and selective inhibitors of the mammalian target of rapamycin. *Cancer Res.* 69, 6232–6240.
- (14) Feldman, M. E., Apsel, B., Uotila, A., Loewith, R., Knight, Z. A., Ruggero, D., and Shokat, K. M. (2009) Active-site inhibitors of mTOR target rapamycin-resistant outputs of mTORC1 and mTORC2. *PLoS Biol.* 7, e38.
- (15) Heitman, J., Movva, N. R., and Hall, M. N. (1991) Targets for cell cycle arrest by the immunosuppressant rapamycin in yeast. *Science* 253, 905–909.
- (16) Kunz, J., Henriquez, R., Schneider, U., Deuter-Reinhard, M., Movva, N. R., and Hall, M. N. (1993) Target of rapamycin in yeast, TOR2, is an essential phosphatidylinositol kinase homolog required for G1 progression. *Cell* 73, 585–596.
- (17) Helliwell, S. B., Wagner, P., Kunz, J., Deuter-Reinhard, M., Henriquez, R., and Hall, M. N. (1994) TOR1 and TOR2 are structurally and functionally similar but not identical phosphatidylinositol kinase homologues in yeast. *Mol. Biol. Cell* 5, 105–118.
- (18) Loewith, R., Jacinto, E., Wullschleger, S., Lorberg, A., Crespo, J. L., Bonenfant, D., Oppliger, W., Jenoe, P., and Hall, M. N. (2002) Two TOR complexes, only one of which is rapamycin sensitive, have distinct roles in cell growth control. *Mol. Cell* 10, 457–468.
- (19) Komeili, A., Wedaman, K. P., O’Shea, E. K., and Powers, T. (2000) Mechanism of metabolic control. Target of rapamycin signaling links nitrogen quality to the activity of the Rtg1 and Rtg3 transcription factors. *J. Cell Biol.* 151, 863–878.
- (20) Mulet, J. M., Martin, D. E., Loewith, R., and Hall, M. N. (2006) Mutual antagonism of target of rapamycin and calcineurin signaling. *J. Biol. Chem.* 281, 33000–33007.
- (21) Aronova, S., Wedaman, K., Anderson, S., Yates, J., 3rd, and Powers, T. (2007) Probing the membrane environment of the TOR kinases reveals functional interactions between TORC1, actin, and membrane trafficking in *Saccharomyces cerevisiae*. *Mol. Biol. Cell* 18, 2779–2794.
- (22) Wullschleger, S., Loewith, R., and Hall, M. N. (2006) TOR signaling in growth and metabolism. *Cell.* 124, 471–484.
- (23) Thoreen, C. C., Kang, S. A., Chang, J. W., Liu, Q., Zhang, J., Gao, Y., Reichling, L. J., Sim, T., Sabatini, D. M., and Gray, N. S. (2009) An ATP-competitive mammalian target of rapamycin inhibitor reveals rapamycin-resistant functions of mTORC1. *J. Biol. Chem.* 284, 8023–8032.